

A study on serodiagnosis of scrub typhus in a Teaching Hospital of South India

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Date of receiving: 3/1/2021

Date if peer review: 12/2/2021

Date of acceptance: 10/3/2021

DOI:10.47799/pimr.0902.03

ABSTRACT

Background: crub typhus is caused by *Orientia tsutsugamushi* (rickettsial disease) commonly transmitted by the bite of larval chiggers of trombiculid mites. It has been one of the important causes of febrile illness, especially in south India. The clinical diagnosis is difficult owing to the non-specific presentation. We in the current study tried to evaluate the serodiagnosis of scrub typhus with the Weil Felix test and IgM ELISA.

Methods: This study was conducted in the Department of Microbiology, Prathima Institute of Medical Sciences, Naganoor, Karimnagar. All the sera samples were subjected to the Weil Felix test using *Proteus* OX2, OX19, OX-K strain agglutination test, and subsequently, Scrub typhus IgM ELISA test.

Results: All the samples were subjected to the Weil Felix test n=4(6.06%) were positive for scrub typhus (OXK antigen) n=11(16.67%) were positive for the spotted group of fever (OX2 antigen) and n=10 (15.15%) were positive of typhus group (OX19 antigen). N=5 sera samples were positive for more than one type of antigens. All the n=66 serum samples were subjected to IgM ELISA for scrub typhus. Out of n=66, only two serum samples (3.03%) were positive by IgM ELISA.

Conclusion: Scrub typhus is emerging as an important public health issue. It is one of the important causes of acute febrile illness. Although it is difficult to distinguish scrub typhus based on the clinical symptoms alone a simple test such as Weil Felix was found to be promising in the diagnosis of scrub typhus. ELISA IgM test may be performed additionally in laboratories with adequate facilities. Hence for clinicians, any case with a

fever of unknown origin should arouse suspicion of scrub typhus.

Keywords: Rickettsial disease, *Orientia tsutsugamushi*, Scrub typhus, Weil-Felix test, IgM

ELISA.

Introduction

Scrub typhus is an acute febrile illness caused by *tsutsugamushi* a type of bacteria that belongs to the Rickettsiaceae family named after *Orientia Tsutsugamushi* (in Japanese means dangerous bug). It is a small gram-negative obligate intracellular organism.^[1]It has caused zoonotic bacterial infections across the world especially in the region of Japan, Taiwan, China, and South Korea including Nepal, Pakistan, Papua New Guinea, Australia, and India. The disease was caused by troops during World War II in Assam and West Bengal and also reported in troops in 1965 of IndoPak war. Southern India has seen its resurgence in recent times.^[1, 2] In countries across Asia the seroepidemiological studies suggest the *Orientia tsutsugamushi* infection ranging from 9.3% to 27.9%. The mortality reports vary widely across the world. Median mortality reports were 6.0% for untreated cases and 1.4% for treated cases of scrub typhus. The mortality rates in south India is reported to be slightly higher at 9%.^[3, 4]Trombiculid mites (chiggers) of the *Leptotrombidium* defense group transmit the disease by the bite of its larva they are microscopic often brilliantly red-colored. Infected chiggers are found in areas of heavy scrub vegetations during the wet season hence sometimes referred to as flood fever when mites lay eggs.^[1]the pathology of scrub typhus is focal or

disseminated vasculitis which occurs due to the destruction of endothelial cells and perivascular infiltration of leucocytes causing self-limiting febrile to fulminant sepsis syndrome.^[5] The clinical presentation is fever which in the endemic areas also known as fever of unknown origin. It is typically associated with rash, myalgia, and lymphadenopathy. The necrotic eschar at the inoculating site of mite is pathognomic of scrub typhus it resembles the skin burn of a cigarette butt.^[6] The complications of scrub typhus are evident after a week of illness which includes renal failure, jaundice, pneumonitis, ARDS, Septic shock, myocarditis, and meningoencephalitis.^[6] There are a variety of tests to detect Scrub typhus such as the Weil Felix test, Indirect Immunofluorescence assay test (IFAT), Immunoperoxidase assay, Enzyme-Linked Immunosorbent Assay (ELISA), Immunochromatography or Rapid diagnostic test (RDT), and PCR.^[7] IFAT, ICT, and PCR are not routinely used in India due to their non-availability, requirement of skilled personnel, and high cost. Weil Felix has shown reasonably high specificity but low sensitivity for the diagnosis of Scrub typhus. Despite all the drawbacks associated with it, the Weil Felix test still serves as a useful and cheapest available screening tool for the laboratory diagnosis of Rickettsial diseases.^[7] Weil-Felix test (W-F) using Proteus OX2, OX19, OX-K strain agglutination test is commercially available for serodiagnostic test and is in use for many years. Only 50% of patients will show positive test results during the second week of illness. A positive titer =1:80 or a four-fold rise over previous levels is significant. Scrub typhus-specific IgM ELISA has shown almost equivalent sensitivity and specificity to those of IFA gold standard, and it can be performed by most laboratories because it does not require any special equipment or technical training.⁹

Materials and Methods:

This cross-sectional study was conducted in the Department of Microbiology Prathima Institute of Medical Sciences, Naganoor, Karimnagar. Blood samples from patients of fever of unknown origin were received from General Medicine and Pediatrics OPDs.

Inclusion criteria

1. History of unknown fever for more than 5 days admitted to the hospital.
2. Clinical features are suggestive of Rickettsial infections such as rash, lymphadenopathy, hepatosplenomegaly.

3. Eschar or history of the mite bite

4. Age above 7 years.

Exclusion criteria

1. Patients with other established causes of infections
2. Hemolysed and lipemic sera specimens.

Blood Sample Collection: Under aseptic precautions 5-10ml venous blood were collected from suspected cases of scrub typhus in a sterile vacutainer tube after getting informed consent for a serological test. Blood was allowed to clot at room temperature for half an hour and serum was separated by centrifugation at 3000rpm for 5 minutes. All the serum samples were subjected to the Weil Felix test followed by IgM ELISA as per the manufacturer's instructions

Weil-Felix Test: The Weil-Felix Proteus agglutination assay (P. Vulgaris OX-19, OX-2, & P. mirabilis OX-K strain agglutination), (lab 21 healthcare limited) was performed on each sample according to the manufacturer's instructions by diluting each serum 1/20 to 1/1280. A single Weil-Felix titer of =1: 80 were accepted as positive results.

IgM ELISA Test: IgM ELISA test was performed on each sample as per the manufacturer's instructions provided along with the kit (InBios international. Inc). Optical density (OD) values were recorded in an ELISA reader by using a 450nm filter. More than 0.5 OD value was positive for IgM Scrub typhus in the test sera.

Results

N=66 samples were collected from the patients with acute febrile illness, fulfilled the eligibility criteria were included in the study. Out of n=66 patients n=39 (59.09%) were males and 27 (40.90%) were females. The age group wise 7 – 25 years were n=28 cases 26 – 45 n=22 cases and > 46 n=16 cases. All the samples were subjected to the Weil Felix test n=4 (6.06%) were positive for scrub typhus (OXK antigen) n=11 (16.67%) were positive for the spotted group of fever (OX2 antigen) and n=10 (15.15%) were positive of typhus group (OX19 antigen). N=5 sera samples were positive for more than one type of antigens. All the n=66 serum samples were subjected to IgM ELISA for scrub typhus. Out of n=66, only two serum samples (3.03%) were positive by IgM ELISA.

Table 1: Shows prevalence of Rickettsial infection among the study group.

| Weil-Felix Test Positive | Frequency (N=66) | Percentage |
|--------------------------|------------------|------------|
| OX2 (SPOTTED FEVER SGF) | 6+5* | 16.67% |
| OX19 (TYPHUS GROUP) | 5+5* | 15.15% |
| OXK (SCRUB TYPHUS) | 2+2* | 6.06% |
| OX2* + OX19* | 3 | 4.54% |

| | | |
|--------------|---|-------|
| OX19* + OXK* | 1 | 1.51% |
| OXK* + OX2* | 1 | 1.51% |

Out of 66 samples, 4 (6.06%) were positive for scrub typhus OXK by Weil Felix used as a screening test and, 2 (3.03%)

were found positive by both Weil Felix heterophile agglutination test and the scrub typhus IgM ELISA (Table 2).

Table 2: Prevalence of Scrub typhus among the study group

| Test | Frequency | Percent |
|--------------------------|-----------|---------|
| Weil Felix Test (OXK) | 4 | 6.06% |
| IgM ELISA (Scrub Typhus) | 2 | 3.03% |

Out of n=4 scrub typhus positive by Weil Felix test OXK, 3 (75%) were male and 1 (25%) were female. Out of n=11 positives for a spotted group of fever by Weil Felix test OX2, 8 (72.72%) were male and n=3 (27.27%) were female. Out of n=10 positives for typhus group OX19, 7 (70%) were male and 3 (30%) were females. Table 3 shows that the prevalence of rickettsial

infection (scrub typhus) is high in between 7-25 years of age group, (scrub typhus 50%, spotted fever 54.54%, and typhus fever group 50.0%). But the finding is not statistically significant (p-value >0.05).

Table 3: Prevalence of scrub typhus among the different age groups.

| Weil Felix test | Frequency of positive | Age in years | | | P-values |
|-----------------|-----------------------|--------------|------------|------------|----------|
| | | 7 – 25yrs | 26 - 45yrs | >46yrs | |
| OXK | 4 | 2 (50%) | 1 (25%) | 1 (25%) | 0.22 |
| OX2 | 11 | 6 (54.54%) | 2 (18.18%) | 3 (27.27%) | 0.146 |
| OX19 | 10 | 5 (50%) | 2 (20%) | 3 (30%) | 0.339 |

Out of 4 patients showing positive agglutination of significant titer for scrub typhus by Weil Felix test, 75% of patients had a headache, arthralgia, and myalgia, 50% of patients had chills, 25% of patients had maculopapular rashes

and hepatomegaly seen in 25% patients and the mean duration of the fever were 11 days (7 to 20 days).

Table 4: common presentation in scrub typhus patients

| Clinical presentation of Scrub typhus (n=4) | Percentage |
|---|------------|
| Headache | 75 |
| Arthralgia | 75 |
| Myalgia | 75 |
| Chills | 50 |
| Rashes (Maculopapular) | 25 |
| Nausea & Vomiting | 25 |
| Hepatomegaly | 25 |

Discussion

In this study, we used single acute-phase sera from n=66 patients of which n=39 (59.09%) were males and n=27 (40.90%) were females. 49 (58.3%) with acute febrile illness attending the hospital for treatment and determining antibodies against

SFG, TG, and ST. With the Weil Felix test n=4 (6.06%) were positive for scrub typhus (OXK antigen) n=11 (16.67%) were positive for the spotted group of fever (OX2 antigen) and n=10 (15.15%) were positive of typhus group (OX19 antigen). A study by Kulkarni et al;^[8] from the Western part of India reported a higher incidence of spotted fever group. Our study also showed

more positives for the spotted fever group followed by typhus fever and scrub typhus. Rathi et al;^[9] also reported that of the n=75 patients with Rickettsial infections, n=52 (69.3%) had spotted fever and n=23 (30.7%) scrub typhus n=56. Mittal et al;^[10] conducting a study on the fever of unknown origin found 42.6% were positive for OXK, 39.3% were positive for OX2 and 8.1% were positive for OX19. All the samples when subjected to IgM ELISA scrub typhus found n=2(3.03%) were positive for scrub typhus with OD values of 0.5. Indirect IgM ELSA may give false-positive results due to rheumatoid factor and false-negative due to an increase of IgG titers at the time of secondary infection.^[11] The primary infection produces a rapid rise in IgM antibodies within 8 days, whereas secondary or re-infection is characterized by a sharp rise in IgG levels, with a variable IgM response.^[12] In this study, the prevalence of scrub typhus was 6.06% by using Weil Felix heterophile agglutination test used as a screening test and 3.03% by IgM ELISA for scrub typhus. There are varying reports of the prevalence of scrub typhus in different parts of India. It is especially true in the case of tertiary care hospitals as the majority of patients seek health care in primary care hospitals.^[7] The transmission of disease occurs throughout the year in tropical areas, in temperate zones, the transmission is seasonal the seasonality of the disease is determined by the appearance of larvae which is found mainly in autumn and spring seasons^[13] In this study we found most of the cases reported between November and March. Mathai et al;^[14] have reported an outbreak of scrub typhus in the winter months. We found male preponderance in this study. On the contrary Sharma et al;^[15] have found a greater number of cases of scrub typhus in females because they commonly worked in fields. The clinical presentation is generally of acute febrile illness with no specific signs and symptoms.^[3] In the past, the clinical diagnosis of scrub typhus was depended on the detection of eschar and rash with a history of outdoor activity.^[16] AR Chogle, et al;^[7] said that the presence of eschar is an important finding for the diagnosis of Rickettsialpox, cutaneous anthrax, tick-borne Rickettsiosis, and other diseases. Although eschars have a high diagnostic value, the lesions are painless and without any itching sensation in most cases, causing the infection to be undetected by most patients. The test in current use is the Weil-Felix OX-K agglutination reaction, which is inexpensive, easy to perform, and results are available overnight; however, it lacks specificity and sensitivity.^[7] For the initial diagnosis of scrub typhus in the present study, the Weil-Felix test was used. This test showed more positives when compared with ELISA and N-PCR tests. It was also seen that there was good agreement between the Weil-Felix test and ELISA when compared with N-PCR. Hence, the Weil-Felix test and ELISA tests can be used in laboratories where PCR is not available. PCR methods when used independently or in conjugation with the Weil-Felix test can be employed as a specific diagnostic tool for the diagnosis of scrub typhus in developing countries and aid in the surveillance

and effective treatment of this emerging infectious disease.^[17] A commercially available ELISA for immunoglobulin M (IgM) and IgG detection using r56 has been developed and evaluated previously. The r56 IgM assay may be even more sensitive to differences in immune responses to the infecting strains than the IIP or the MIF assay because no other conserved antigens are present as found in whole-organism assays.^[18] The ELISA format is very convenient for large-scale testing in the laboratory.^[19] The serological tests have low sensitivities in the early stage of scrub typhus due to insufficient production of antibodies, frequent follow-up tests are needed.^[20] Thus, a rapid early and accurate diagnosis of scrub typhus is essential for specific and effective treatment

Conclusion

Within the limitations of the current study, we found Scrub typhus is emerging as an important public health issue. It is one of the important causes of acute febrile illness. Although it is difficult to distinguish scrub typhus based on the clinical symptoms alone a simple test such as Weil Felix was found to be promising in the diagnosis of scrub typhus. ELISA IgM test may be performed additionally in laboratories with adequate facilities. Hence for clinicians, any case with a fever of unknown origin should arouse suspicion of scrub typhus.

REFERENCES

1. Rapsang AG, Bhattacharyya P. Scrub typhus. *Indian J Anaesth* 2013;57:127-34.
2. Chakraborty S, Sarma N. Scrub typhus: An emerging threat. *Indian J Dermatol* 2017;62:478-85
3. Bonell A, Y Lubell, PN Newton, JA Crump, DH Paris. Estimating the burden of scrub typhus: A systematic review. *PLoS Negl Trop Dis* 2017;11(9): e0005838
4. Paul Trowbridge, Divya P, PS Premkumar, GM Varghese. Prevalence and risk factors for scrub typhus in South India. *Tropical Medicine and International Health* 2017; 22(5):576–582.
5. Anugrah Chrispal, H Boorugu, KG Gopinath, JAS Jude Prakash. Scrub typhus: an unrecognized threat in South India clinical profile and predictors of mortality. *tropical doctor* 2010; 40: 129–133.
6. SK Mahajan. Scrub Typhus. *JAPI* 2010;53:954-58.
7. AR Chogle. Diagnosis and Treatment of Scrub Typhus - The Indian Scenario. *J Assoc Physician India* 2010; 58: 11-12.
8. Scrub Typhus - Centers for Disease Control and Prevention (2017) <https://www.cdc.gov/typhus/scrub/index.html> [Last accessed on 20 Nov 2020]

9. Kulkarni A, et al. Rickettsial disease- an expensive. *Pediatric infect Dis* 2009;1:118-124.
10. Rathi NB, Rathi AN, Goodman MH, Aghai ZH. Rickettsial diseases in central India: proposed clinical scoring system for early detection of spotted fever. *Indian Pediatr* 2011; 48(11):867-72.
11. Jang WJ, et al. Evaluation of an immunoglobulin M capture enzyme linked immunosorbent assay for diagnosis of *Orientia tsutsugamushi* infection. *Clin Diagn Lab Immunol* 2003;10:394-8.
12. D J Kelly, PA Fueerst, Wei-Mei Ching, A L Richards. Scrub typhus: The geographic distribution of phenotypic and genotypic variants of *Orientia tsutsugamushi*. *Clin Infect Dis* 2009. 48 (3):203–30.
13. Raoult D. *Orientia tsutsugamushi*. In: Mandell GL, Bennett JE, Dolin R (eds). *Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases*. 7th ed. Philadelphia: Churchill Livingstone; 2009. Pp. 2529-30.
14. Mathai E, Rolain JM, Verghese GM, Abraham OC, Mathai D, Mathai M, Raoult D. Outbreak of scrub typhus in southern India during the cooler months. *Ann N Acad Sci* 2003; 990:359-64.
15. Sharma A, Mahajan S, Gupta ML, Kanga A, Sharma V. Investigation of an outbreak of scrub typhus in the Himalayan region of India. *Jpn J Infect Dis* 2005; 58: 208-20.
16. Blake FG, Maxcy KF, Sadusk JF, Kohls GM, Bell EJ. Studies on *tsutsugamushi* disease (scrub typhus, mite-borne typhus) in New Guinea and adjacent islands: epidemiology, clinical observations, and etiology in the Dobadura. *Am J Hyg* 1945(b); 41:243-373.
17. Blacksell SD, Bryant NJ, Paris DH, Doust JA, Sakoda Y, Day NP. Scrub typhus serologic testing with the indirect immunofluorescence method as a diagnostic gold standard: a lack of consensus leads to a lot of confusion. *Clin Infect Dis* 2007; 44(3):391-401.
18. Land MV, Ching WM, Dasch GA, Zhang Z, Kelly DJ, Graves SR, Devine PL. Evaluation of a commercially available recombinant-protein enzyme-linked immunosorbent assay for detection of antibodies produced in scrub typhus rickettsial infections. *J Clin Microbiol* 2000; 38(7):2701-5.
19. Ching WM, et al. Early diagnosis of scrub typhus with a rapid flow assay using recombinant major outer membrane protein antigen (r56) of *Orientia tsutsugamushi*. *Clin Diagn Lab Immunol* 2001; 8(2):409-14.
20. Bozeman FM, Elisberg BL. Serological diagnosis of scrub typhus by indirect immunofluorescence. *Proc Soc Exp Biol Med* 1963; 112: 568-73.

How to cite this article : Kelamane S, Cheruku Pranuthi Mispah, K. Sri Sandhya. A study on serodiagnosis of scrub typhus in a Teaching Hospital of South India. *Perspectives in Medical Research* 2021; 9 (2):10-14

DOI:10.47799/pimr.0902.03

Sources of Support: Nil, Conflict of interest: None declared